



ZR-96 Oligo Clean & Concentrator™

Clean-up DNA/RNA oligos from any reaction

Highlights

- High-throughput (96-well), spin-plate clean-up of ultra-pure DNA and RNA oligonucleotides.
- Complete removal of dyes, salts, enzymes, nucleotides and short ٠ oligos.
- Eluted DNA/RNA (≥ 10 µl) is ready for hybridization, sequencing, PCR, ligation, etc.

Catalog Numbers: D4062, D4063



Scan with your smart-phone camera to view the online protocol/video.







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Product Contents

ZR-96 Oligo Clean & Concentrator™	D4062 (2 x 96 prep)	D4063 (4 x 96 prep)
Oligo Binding Buffer	40 ml	40 ml (x2)
DNA Wash Buffer (concentrate) ¹	48 ml	48 ml (x2)
Zymo-Spin [™] I-96 Plate	2	4
Collection Plate	4	8
Elution Plate	2	4
96-Well Cover Foil	2	4
Instruction Manual	1	1

Storage Temperature - Store all kit components (i.e., buffers, columns) at room temperature.

Before use:

1 Prior to use, add 192 ml 100% ethanol (208 ml of 95% ethanol) to the 48 ml DNA Wash Buffer concentrate .

Specifications

- **Sample Sources** Enzymatic reactions mixtures containing oligonucleotides ≥ 16 nt (radioactive-, biotin-, DIG-labeled, etc.)
- Size Limits For oligonucleotides \geq 16 nt, up to 23 kb.
- **Purity** A₂₆₀/A₂₈₀ & A₂₆₀/A₂₃₀ > 1.8. DNA/RNA is ready for hybridization, sequencing, ligation, PCR and etc.
- **Binding Capacity** 10 µg ssDNA/RNA or 5 µg dsDNA per well, with a typical recovery of > 90% (**Zymo-Spin**[™] **I-96 Plate**).
- Elution Volume \geq 10 µl (water not provided).
- Detergent Tolerance ≤ 5% Triton X-100, ≤ 5% Tween-20, ≤ 5% Sarkosyl, ≤ 0.1% SDS.
- Equipment Needed (user provided) Centrifuge with a spinplate rotor.

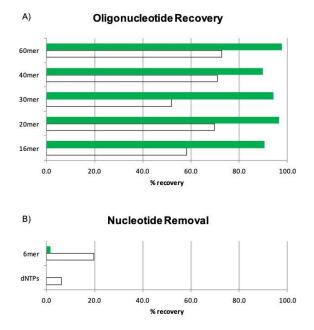
Applications

- Isotope and Dye Removal Efficiently removes unincorporated fluorescent (i.e., AMCA, FITC, BIO, DIG, Cy3, Cy5, FAM, etc.) and radiolabeled dNTP derivatives from DNA following in vitro labeling reactions.
- DNA Fragment Clean-up from Enzymatic Reactions Desalting of DNA with the removal of DNA polymerases, modifying enzymes, RNA polymerases, ligases, kinases, nucleases, phosphatases, endonucleases, etc.
- Post-Reverse Transcription (RT) and cDNA Clean-up Purifies DNA following RT, either as a DNA/RNA complex or as single stranded cDNA following chemical hydrolysis of the RNA template (page 6).

Product Description

The **ZR-96 Oligo Clean & Concentrator**^{$^{\text{M}}$} kit provides a streamlined method for high-throughput (96-well), efficient recovery and clean-up of DNA and RNA oligonucleotides \geq 16 nt from labeling (radioactive, biotin, DIG, *etc.*) and other enzymatic reactions. Unincorporated nucleotides, short oligos, dyes, enzymes, and salts are effectively removed by the clean-up procedure.

There is no need for organic denaturants or chloroform. Instead, the kit features Zymo-Spin[™] plate technology and employs a single-buffer system that allows for efficient oligonucleotide adsorption to the matrix of each well in the **Zymo-Spin[™]I-96 Plate**. Oligonucleotide is washed and concentrated into a small volume of water ($\geq 10 \ \mu$ I). Purified oligonucleotide is suitable for hybridization, gel shift assays, enzymatic reactions, ligation, sequencing, microarray analysis, *etc*.



Oligo Clean & Concentrator[™] facilitates > 90% recovery of ssDNA oligonucleotides (A) and efficient short oligo and nucleotide removal (B).

Protocol

The protocol consists of: (I) Buffer Preparation and (II) DNA/RNA Clean-Up

(I) Buffer Preparation

 Prior to use, add 192 ml 100% ethanol (208 ml of 95% ethanol) to the 48 ml DNA Wash Buffer concentrate.

(II) DNA/RNA Clean-up

- ✓ Perform all steps at room temperature and centrifugation at ≥ 2,500 x g for 5 minutes, unless specified.
- ✓ Scale up the volumes proportionally (steps 1-2), if needed.
- ✓ Do not use the **96-Well Plate Cover Foil** on the spin-plate during DNA/RNA Clean-Up. If necessary, use an Air Permeable Sealing Cover (#C2011-8); sold separately.
- 1. Add 100 µl Oligo Binding Buffer to 50 µl sample¹.
- 2. Add 400 µl ethanol² (95-100%) and mix well by pipetting.
- 3. Transfer the sample to each well of the **Zymo-Spin[™] I-96 Plate**³ mounted on a **Collection Plate** and centrifuge. Discard the flow-through⁴.
- Add 750 µl DNA Wash Buffer to the column and centrifuge. Then centrifuge again to ensure complete removal of the wash buffer. Carefully, transfer the plate onto an Elution Plate.
- 5. Add 25 µl water⁵ directly to the matrix of each well and centrifuge.

Alternatively, for highly concentrated DNA/RNA use \geq 10 µl elution.

The eluted oligonucleotide can be used immediately or stored frozen. Use the **96-Well Cover Foil** to prevent the eluate from evaporation.

¹ To minimize pipetting error, adjust the sample volume to 50 µl (minimum).

² For DNA/RNA \geq 80 nt, only 200 μl ethanol is required.

³ To process samples >700 µl, Zymo-Spin[™] plate may be reloaded.

⁴ For **radioactive samples**, transfer the column into a new **Collection Tube** and discard the tube containing the radioactive flow-through appropriately.

⁵ Alternatively, TE buffer can be used for elution (if required).

Appendices

cDNA Clean-Up following Reverse Transcription (RT)

The **Oligo Clean & Concentrator** can be used to effectively clean and concentrate first-strand cDNA following reverse transcription (RT) and hydrolysis. The **Oligo Binding Buffer** will neutralize the hydrolysis reaction and the recovered cDNA may be used directly for microarray analysis. etc.

Hydrolysis Reaction: To each 30-50 μ I RT reaction, add 10 μ I 0.5 M EDTA and 10 μ I 1 M NaOH. Then mix and incubate at 65°C for 15 minutes. Proceed to the DNA/RNA Clean-Up protocol, page 5.

Format Compatibility

To adjust binding capacity, simply replace the provided columns with the formats indicated below and follow their respective elution volumes.

Format	Zymo-Spin™ I & IC	Zymo-Spin™ II & IICR	Zymo-Spin™ III & IIICG	Zymo-Spin™ I-96 Plate
Item Image		Ţ		
Kit Name	<u>OCC™</u>	N/A	N/A	<u>ZR-96 OCC™</u>
Capacity	10 μg ssDNA/RNA or 5 μg dsDNA / prep.	50 μg ssDNA/RNA or 25 μg dsDNA / prep.	100 μg ssDNA/RNA or 50 μg dsDNA / prep.	10 μg ssDNA/RNA or 5 μg dsDNA / prep.
Elution Vol.	≥ 6 µl	≥ 25 µl	≥ 50 µl	≥ 10 µl
Column Cat. Nos.	<u>C1003-50, C1004-50</u>	<u>C1008-50, C1078-50</u>	<u>C1005-50, C1006-50-G</u>	<u>C2004</u>

Ordering Information

Product Description	Catalog No.	Size
ZR-96 Oligo Clean & Concentrator™	D4062 D4063	2 x 96 preps. 4 x 96 preps.

Individual Kit Components	Catalog No.	Amount
Oligo Binding Buffer	D4060-1-10 D4060-1-40	10 ml 40 ml
DNA Wash Buffer (concentrate)	D4003-2-24 D4003-2-48	24 ml 48 ml
Zymo-Spin™ I-96 Plate	C2004	2 plates
Collection Plate	C2002	2 plates
Elution Plate	C2003	2 plates

Complete Your Workflow

✓ For tough-to-lyse samples in TRIzol, use ZR BashingBead Lysis Tubes:

ZR BashingBead Lysis Tubes	
2.0 mm beads #S6003	For plant/animal tissue
0.1 + 0.5 mm beads #S6012	For microbes
0.1 + 2.0 mm beads #S6014	For microbes in tissue/insects

✓ The only direct, high-throughput and automatable RNA purification from sample lysates in TRIzol (DNase I Set included with all formats):



Direct-zol RNA kits	
Microprep #R2060-R2063	From 1 cell and up
Miniprep #R2050-R2053	Up to 50 ug RNA
Miniprep Plus #R2070-R2073	Up to 100 ug RNA
96-well #R2054-R2057	Spin-plate
MagBeads #R2100-R2105	Automatable (Tecan, Hamilton, Kingfisher, etc.)

✓ For RNA clean-up (purification) from the aqueous phase (e.g., TRIzol, TRI Reagent or similar) or from any enzymatic reaction (e.g., DNase I treated RNA):



RNA Clean & Concentrator kit

#R1013-R1014

DNase I Set included

✓ For NGS:

Zymo-Seq RiboFree Total RNA Library Prep kit		
#R3000	12 preps	
#R3003	96 preps	



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Integrity of kit components is guaranteed for up to one year from date of purchase. Reagents are routinely tested on a lot-to-lot basis to ensure they provide the highest performance and reliability.

This product is for research use only and should only be used by trained professionals. It is not for use in diagnostic procedures. Some reagents included with this kit are irritants. Wear protective gloves and eye protection. Follow the safety guidelines and rules enacted by your research institution or facility.

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The **BEAUTY** of **SCIENCE** is to Make Things \textbf{SIMPLE}°