ZYMO RESEARCH

DNA Purification Made Simple

Quick-DNA™ Microprep Plus Kit

Purify high quality total DNA from a variety of sample types.

Highlights

- Extract high-quality DNA easily and reliably from any biological fluids, cultured/monolayer cells, or solid tissues.
- **Zymo-Spin™ Technology** ensures DNA is ready for all sensitive downstream applications such as qPCR, DNA-sequencing, arrays, and methylation analysis.

Catalog Numbers:
D4074

Scan with your smart-phone camera to view the online protocol/video.

tech@zymoresearch.com www.zymoresearch.com Toll Free: (888) 882-9682

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Product Contents

Quick-DNA™ Microprep Plus Kit	D4074 (50 Preps.)	Storage Temperature
Proteinase K & Storage Buffer	2 x 5 mg	−20°C (after mixing)
BioFluid & Cell Buffer (Red)	6 ml	Room Temp.
Solid Tissue Buffer (Blue)[1]	6 ml	Room Temp.
Genomic Binding Buffer	25 ml	Room Temp.
DNA Pre-Wash Buffer[1]	15 ml	Room Temp.
g-DNA Wash Buffer	50 ml	Room Temp.
DNA Elution Buffer	10 ml	Room Temp.
Zymo-Spin™ IC-XM Columns	50 Columns	Room Temp.
Collection Tubes	100 Tubes	Room Temp.
Instruction Manual	1	-

[1] The **Solid Tissue Buffer (Blue)** and **DNA Pre-Wash Buffer** may have formed a precipitate. If this is the case, incubate at 37°C to solubilize. DO NOT MICROWAVE.

Specifications

- **Sample Sources** – See pages 3 and 4.
- **Workflow Overview** – Utilizes a Proteinase K Digestion and Zymo-Spin™ Technology for effective recovery of DNA. See page 6 for more information.
- **DNA Types** – The ***Quick*-DNA™ Microprep Plus Kit** will isolate total DNA including genomic, mitochondrial, plasmid, viral, parasitic, etc. from biological fluids, cells, or tissues. Not recommended for small cell-free DNA isolation from urine, serum, and plasma.
- **DNA Purity** – High quality DNA is ready for all sensitive downstream applications such as PCR, endonuclease digestion, Southern blotting, genotyping, Next-Generation Sequencing, bisulfite conversion, etc. ($A_{260}/A_{230} \geq 2.0$).
- **DNA Size** – Capable of recovering genomic and mitochondrial DNA sized fragments > 50 kb. If present, parasitic, microbial, and viral DNA will also be recovered.
- **DNA Yield** – The DNA binding capacity of each column is 5 µg. Typically, mammalian tissues yield: 1-3 µg DNA per mg skeletal, heart, lung, and brain tissues and 3-5 µg DNA per mg liver and kidney. Human whole blood will yield 3-7 µg DNA per 100 µl blood sampled.
- **Elution Volume** – DNA can be eluted into as little as 10 µl **DNA Elution Buffer** or water**.**
- **Equipment** – Water bath or heat block (55˚C), microcentrifuge, and vortex.
- **DNA Applications** – DNA isolated using the ***Quick*-DNA™ Microprep Plus Kit** can be used for life-science research, genotyping, livestock breeding, veterinary research, and routine applied testing among a variety of other applications.

Sample Sources

Biological Fluids: For total DNA isolation from ≤ 50 µl of whole blood, nucleated blood, buffy coat, saliva, sputum, semen, milk, etc.

Special Considerations:

- For biological fluids samples stored in DNA/RNA Shield™, see **Samples in DNA/RNA Shield™** (**pg. 11**).
- For nucleated blood samples, such as avian blood, see **Nucleated Blood Samples** (**pg. 12**).
- For blood, saliva, and cells collected onto Guthrie, FTA®, and other storage papers (cards), see **Samples Collected onto Storage Papers/Cards** (**pg. 16**)
- For viral DNA isolation from serum/plasma samples, follow the Biological Fluids & Cells workflow. Not recommended for small cell-free DNA isolation from serum/plasma. For small cell-free DNA isolation from serum/plasma samples, use the ***Quick*-cfDNA™ Serum & Plasma Kit (D4076).**
- To isolate cellular and/or cell-free DNA from up to 40 ml of urine samples, see the ***Quick*-DNA™ Urine Kit (D3061)**. For cellular DNA from urine, pellet at 3,000 x *g* for 15 minutes and remove supernatant before processing using the Biological Fluids & Cells workflow.

Mammalian/Insect Cell Cultures: For total DNA isolation from ≤ 1 x 10^6 cells such as HeLa cells, HEK-293 cells, *Drosophila* cell lines, etc.

Special Considerations:

- Media should be removed before processing by pelleting cells (at approximately 500 x *g* for 2 minutes depending on volume and cell type) and removing the supernatant.
- For mammalian cell samples, it is possible to reduce Proteinase K digestion time to 5 minutes at 55˚C (Step 2 on **pg. 7**).
- For cell monolayer and buccal cell preparation and collection, see **Cell Monolayer Sample Preparation (pg. 9** and **10)**.
- For samples stored in DNA/RNA Shield™, see **Samples in DNA/RNA Shield** (**pg. 11**).

Sample Sources (continued)

Bacterial Cell Cultures: For total DNA isolation (e.g., genomic, plasmid, etc.) from ≤ 1 x 10^6 *E. coli* cells.

Special Considerations:

- Media should be removed before processing by pelleting cells (pellet cells at approximately 500 x *g* for 2 minutes depending on volume and cell type) and removing the supernatant.
- For *E. coli* samples and other easy to lyse microbes, follow the Biological Fluids & Cells workflow. All other bacterial samples may be resistant to chemical lysis and Proteinase K digestion and should be used with the ***Quick*-DNA™ Fungal/Bacterial DNA Microprep Kit** (D6007).
- Microbes previously lysed with enzymes (e.g. Lysozyme) or other mechanical methods (e.g. bead beating or liquid nitrogen) may be processed by using the Biological Fluids & Cells workflow.

Solid Tissues: For total DNA isolation from ≤ 5 mg tail snips, ear punches, organ biopsies (brain, liver, heart, kidney, muscle, stomach, bladder, intestine, etc.).

Special Considerations:

- Overnight Proteinase K digestion at 55˚C is possible (Step 2, **pg. 7**).
- For solid tissue samples stored in DNA/RNA Shield™, see Samples in DNA/RNA Shield (**pg. 11**).
- For hair, fingernail, feather, and bone samples see **Hair, Fingernail, Feather, and Bone Samples (pg.13).**
- For FFPE samples, see *Quick*-DNA™ FFPE Kit (D3067) for specialized FFPE DNA purification. See **FFPE Samples** (**pg. 14**) for an adapted protocol using the *Quick*-DNA™ Microprep Plus Kit.

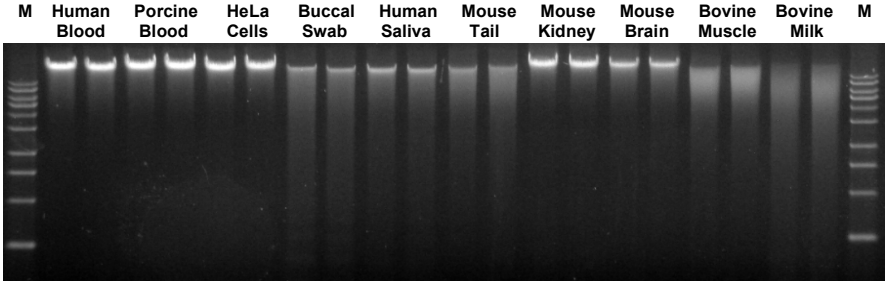
Tough-to-Lyse Samples: For total DNA isolation from fungal, bacterial, plant/seed, insect, fecal, and soil samples.

Special Considerations:

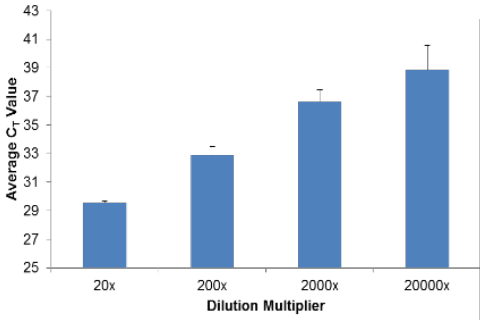
- **Microbiomics and Metagenomics:** Use the **ZymoBIOMICS® DNA Microprep Kit** (D4301) for accurate community profiling. The **ZymoBIOMICS® DNA Microprep Kit** also includes innovative inhibitor removal technology enabling purification of inhibitor free DNA from nearly any sample type (feces, soil, water, biofilms etc.).
- **Microbial Isolation from Environmental Samples:** For samples not intended for community profiling, use the ***Quick*-DNA™ Fecal/Soil Microbe Microprep Kit** (D6012).
- **Plants and Seeds:** Use the ***Quick*-DNA™ Plant/Seed Miniprep Kit** (D6020).

Product Description

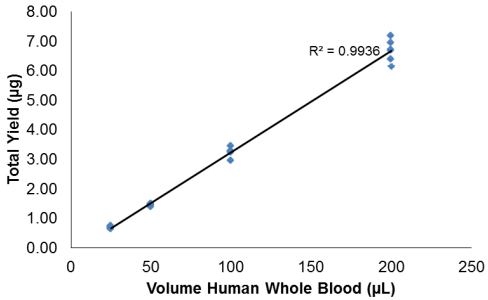
The ***Quick*-DNA™ Microprep Plus Kit** is the easiest method for high yield total DNA extraction (e.g., genomic, mitochondrial, viral) from any biological fluid, cell culture, or solid tissue sample. Innovative reagents and Zymo-Spin Technology allow for ultra-pure and concentrated genomic DNA > 50 kb to be eluted in as little as 10 µl. Zymo-Spin™ Columns ensure no buffer retention. Purified DNA is RNA-free bypassing the need for RNase A treatment and ensuring accurate quantification for applications like library preparations. Isolated DNA is suitable for immediate use in sensitive downstream applications including qPCR, DNA-seq, arrays, and methylation analysis.



High Quality DNA Obtained from a Wide Range of Biological Samples Using the *Quick*-DNA™ Miniprep Plus Kit. DNA purified using the *Quick*-DNA™ Miniprep Plus Kit is ultrapure, highly concentrated, and ready for all downstream applications. Input DNA was standardized to 300 ng and analyzed in a 1% (w/v) TAE/agarose/EtBr gel (shown above). The size marker "M" is a 1 kb ladder (Zymo Research).



HSV-1 Viral DNA is Effectively Isolated from Plasma Using the *Quick*-DNA™ Miniprep Plus Kit. A dilution series of HSV-1 spiked into porcine plasma and extracted using the *Quick*-DNA™ Miniprep Plus Kit shows effective purification and subsequent qPCR amplification, even at a 20,000:1 dilution. The no template controls did not amplify even after 50 cycles.

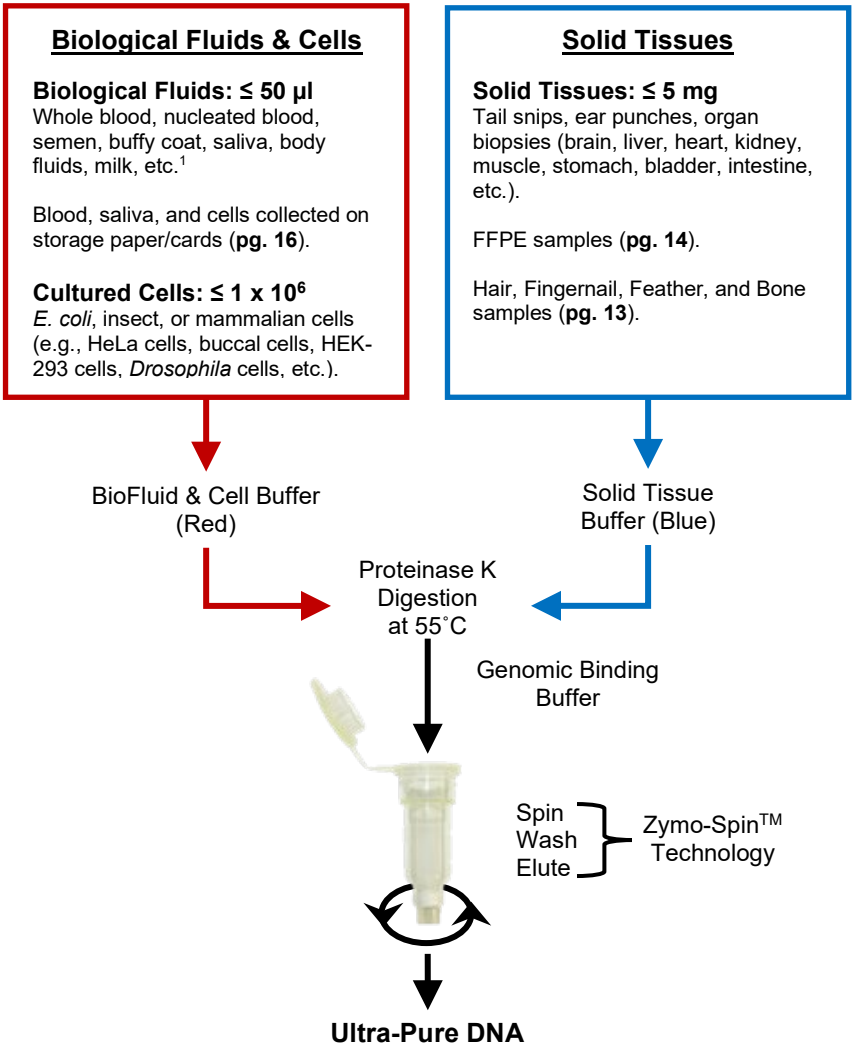


DNA Yields Increase Linearly with Increasing Volumes of Human Whole Blood Using the *Quick*-DNA™ Miniprep Plus Kit. Six replicates of 25, 50, 100, and 200 µl of human whole blood were processed.

Purification Guide

The ***Quick*-DNA™ Microprep Plus Kit** facilitates high-throughput purification of DNA from any biological fluids, cultured/monolayer cells, or solid tissues by combining enzymatic and chemical extraction regimens.

Quick-DNA™ Microprep Plus Kit Workflow



[1] Viral DNA from serum or plasma samples can also be processed using this workflow. Not recommended for cell-free DNA isolation from urine, serum, or plasma samples. For cell-free DNA isolation from up to 40 ml urine, see the ***Quick*-DNA™ Urine Kit** (D3061). For cell-free DNA isolation from up to 10 ml serum or plasma samples, see the ***Quick*-cfDNA™ Serum & Plasma Kit** (D4076).

Protocol

Reagent Preparation

✓ Add 260 µl **Proteinase K Storage Buffer** to each **Proteinase K** (5 mg) tube prior to use. The final concentration of **Proteinase K** is ~20 mg/ml. Store at -20ºC after mixing.

Sample Processing

*Resuspend ≤ 1 x 10^6 mammalian or insect cell pellets using 50 µl **DNA Elution Buffer** or an isotonic buffer (e.g. PBS).*

Biological Fluids & Cells

1. Add up to 50 µl[1] sample to a microcentrifuge tube and add:

   50 µl **BioFluid & Cell Buffer (Red)**
   5 µl **Proteinase K**

2. Mix thoroughly or vortex 10-15 seconds and then incubate the tube at 55˚C for 10 minutes[2,3].

3. Add 1 volume **Genomic Binding Buffer** to the digested sample. Mix thoroughly or vortex 10-15 seconds.

   Example: Add 105 µl Genomic Binding Buffer to the 105 µl digested sample.

Solid Tissues

1. To tissue samples (≤ 5 mg) in a microcentrifuge tube, add a solution of:

   45 µl Water
   45 µl **Solid Tissue Buffer (Blue)**
   10 µl **Proteinase K**

2. Mix thoroughly or vortex 10-15 seconds and then incubate the tube at 55˚C for 1-3 hours or until tissue solubilizes[2]. Mix thoroughly before proceeding[3].

3. Add 2 volumes **Genomic Binding Buffer** to the supernatant. Mix thoroughly or vortex 10-15 seconds.

   Example: Add 200 µl Genomic Binding Buffer to the 100 µl mixture.

4. Transfer the lysate to the **Zymo-Spin™ IC-XM Column** in a **Collection Tube**. Centrifuge at ≥ 12,000 *x g* for 5 minutes[4]. Discard the flow-through and collection tube.

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[1] If using < 50 µl sample, increase the volume to 50 µl using **DNA Elution Buffer** or an isotonic buffer (e.g. PBS) before continuing.
[2] Overnight digestions are possible without affecting the integrity of the DNA.
[3] To remove insoluble debris, centrifuge at ≥ 12,000 x *g* for 1 minute. Transfer aqueous supernatant to a clean microcentrifuge tube. Avoid transferring the lipid layer and pelleted cellular debris.
[4] If the lysate is still visible on top of the matrix, centrifuge for another minute or until completely cleared.

5. Add 200 µl **DNA Pre-Wash Buffer** to the spin column in a new **Collection Tube**. Centrifuge at ≥ 12,000 x *g* for 1 minute. Empty the Collection Tube.

6. Add 700 µl **g-DNA Wash Buffer** to the spin column. Centrifuge at ≥ 12,000 x *g* for 1 minute. Empty the Collection Tube.

7. Add 200 µl **g-DNA Wash Buffer** to the spin column. Centrifuge at ≥ 12,000 x *g* for 1 minute. Discard the Collection Tube with the flow through.

8. Transfer the spin column to a clean microcentrifuge tube. Add ≥ 10 µl **DNA Elution Buffer** or water[1] directly on the matrix. Incubate for 5 minutes at room temperature, then centrifuge at maximum speed for 1 minute to elute the DNA[2]. The eluted DNA can be used immediately for molecular based applications or stored ≤ -20ºC for future use.

[1] **DNA Elution Buffer**: 10 mM Tris-HCl, pH 8.5, 0.1 mM EDTA. If water is used, ensure the pH is > 6.0.
[2] The total yield can be improved by eluting the DNA with 60-70 °C **DNA Elution Buffer**. Also, loading the eluate a second time, incubating for 3 minutes at room temperature, and centrifuging again can also increase total yield.

Appendices

Cell Monolayer Sample Preparation

The following procedure is designed for up to 1 x 10^6 monolayer cells (dilute if necessary for proper cell counts). Although cell types and culture conditions may vary, the protocol will work with high-density growth cells (e.g., HeLa cells) as well as with low-density growth cells (e.g., neuronal cells).

Trypsinize or scrape adherent cells from a culture flask or plate. Centrifuge the suspension at approximately 500 x *g* for 5 minutes. Remove the supernatant and resuspend the cell pellet in 1 ml PBS (Phosphate Buffered Saline) and then transfer suspension to a microcentrifuge tube. Centrifuge the suspension at approximately 500 x *g* for 5 minutes. Discard the supernatant and then follow the Biological Fluids & Cells workflow on Page 7.

Guidelines for Monolayer Cell DNA Isolation:

Cell numbers (growth densities) can vary between different cell types. Table 1 (below) provides an approximation of the cell numbers that can be recovered from different culture containers for "high-density" growth cells like CV1 and HeLa cells.

Table 1: Culture Plate/Flask Growth Area (cm^2) and Cell Number

Culture Container	Well /Flask Surface Area	Cell Number
96-well plate (each well)	0.32-0.6 cm^2	4-5x10^4
24-well plate (each well)	2 cm^2	1-3x10^5
12-well plate (each well)	4 cm^2	4-5x10^5
6-well plate (each well)	9.5 cm^2	0.5-1x10^6
T25 Culture Flask	25 cm^2	2-3x10^6
T75 Culture Flask	75 cm^2	0.6-1x10^7
T175 Culture Flask	175 cm^2	2-3x10^7

Buccal Cells and Swabs:

Buccal cells can be isolated using a rinse- or swab-based isolation method.

A. **Rinse Method**: Vigorously rinse mouth with 10-20 ml of saline solution or mouthwash orally for 30 seconds. The more vigorous the rinsing action, the more cells that will be recovered. Spit the saline into a 50 ml tube and pellet the cells at 1,500 rpm for minutes. Discard the supernatant without disturbing the cell pellet. Then follow from Step 1 of the Biological Fluids & Cells workflow on Page 7.

B. **Swab Isolation Method:** Thoroughly rinse mouth out with water before isolating cells. Brush the inside of the cheek with a buccal swab for 15 seconds (approximately 20 brushes), making sure to cover the entire area of the inner cheek. Rinse the brush into a microcentrifuge tube using a solution of 100 µl of **BioFluid & Cell Buffer (Red)** and 100 µl TE or another isotonic solution. Add 10 µl of **Proteinase K,** mix thoroughly, and incubate at 55˚C for 10 minutes. Then follow from Step 3 of the Biological Fluids & Cells workflow on Page 7 (dilute or remove sample if needed to reduce the cell count).

Samples in DNA/RNA Shield

DNA/RNA Shield™ *ensures nucleic acid stability during sample storage/transport at ambient temperatures. There is no need for refrigeration or specialized equipment.* ***DNA/RNA Shield™*** *effectively lyses cells and inactivates nucleases and infectious agents (virus), and it is compatible with various collection and storage devices (vacutainers, swabs, nasal, buccal, fecal etc.).*

DNA/RNA Shield™ *purchased separately (R1100 or R1200).*

Biological Fluids and Cell Cultures

1. Add 5 µl of Proteinase K to 100 µl of the sample/shield mixture prepared according to the **DNA/RNA Shield™** specifications.

2. Mix thoroughly or vortex 10-15 seconds and then incubate the tube at room temperature for 20 minutes.

3. Continue from Step 3 of the Biological Fluids & Cells Workflow (**pg. 7**).

Solid Tissues

1. For samples prepared according to the **DNA/RNA Shield™** specifications, homogenize the solid tissue sample by bead bashing or other homogenization protocols.

2. Add 0.5 volumes of the Solid Tissue Buffer **(Blue)** and 10 µl of Proteinase K to the lysate.

3. Mix thoroughly or vortex 10-15 seconds and then incubate the tube at 55˚C for 1 – 3 hours[1].

4. To remove insoluble debris, centrifuge at ≥ 12,000 x *g* for 1 minute. Transfer aqueous supernatant to a clean microcentrifuge tube.

5. Add 1 volume **Genomic Binding Buffer** to the digested sample. Mix thoroughly or vortex 10-15 seconds.

6. Continue from Step 4 of the main protocol (**pg. 7**).

[1] Overnight digestion at 55˚C is possible and will increase the effectiveness of digestion and DNA recoveries.

Nucleated Blood Samples

Add up to 5 µl of nucleated blood to the following in a microcentrifuge tube:

BioFluid & Cell Buffer (Red)	50 µl
Proteinase K	5 µl
DNA Elution Buffer (or TE Solution)	50 µl

1. Mix thoroughly by pipetting up and down. Then incubate the tube at 55°C for 20 minutes[1].

2. Add 1 volume of **Genomic Binding Buffer** to the tube and mix thoroughly by pipetting up and down and by vortexing. Ensure the sample is homogenous before continuing[2].

3. Transfer the mixture to a **Zymo-Spin™ IC-XM Column** in a **Collection Tube**. Centrifuge at ≥ 12,000 *x g* for 1 minute[3]. Discard the collection tube with the flow through.

4. Add 200 µl **DNA Pre-Wash** to the spin column in a new **Collection Tube**. Centrifuge at ≥ 12,000 *x g* for 1 minute. Empty the collection tube.

5. Add 700 µl **g-DNA Wash Buffer** to the spin column. Centrifuge at ≥ 12,000 *x g* for 1 minute. Empty the collection tube.

6. Add 200 µl **g-DNA Wash Buffer** to the spin column directly on the matrix. Centrifuge at ≥ 12,000 *x g* for 1 minute. Discard the collection tube with the flow through.

7. Transfer the spin column to a clean microcentrifuge tube. Add ≥ 10 µl **DNA Elution Buffer** or water[4] directly on the matrix. Incubate for 5 minutes at room temperature, then centrifuge at top speed for 1 minute to elute the DNA[5]. The eluted DNA can be used immediately for molecular based applications or stored ≤ -20ºC for future use.

[1] The sample may not be completely homogenous before digesting.
[2] It may be necessary to pipette up and down many times to ensure the sample is homogenous. Vortexing will also help ensure the mixture is homogenous.
[3] If the lysate is still visible on top of the matrix, centrifuge for another minute or until completely cleared.
[4] **DNA Elution Buffer**: 10 mM Tris-HCl, pH 8.5, 0.1 mM EDTA. If water is used, make sure the pH is > 6.0.
[5] The total yield can be improved by eluting the DNA with 60-70 °C **DNA Elution Buffer**. Also, loading the eluate a second time, incubating for 3 minutes at room temperature, and centrifuging again can also increase total yield.

Hair, Fingernail, Feather, and Bone[1] Samples

1. Freshly prepared DTT (dithiolthreitol) (not provided) needs to be added to each microcentrifuge tube containing sample (≤ 5 mg) as follows:

Water	40 µl
Solid Tissue Buffer (Blue)	45 µl
DTT (1 M)	5 µl
Proteinase K	10 µl

2. Mix thoroughly or vortex 10-15 seconds and then incubate the tube at 55°C for 1-3 hours[2].

3. Add 200 µl **Genomic Binding Buffer** to the tube and mix thoroughly by vortexing for 15 seconds. Centrifuge at ≥ 12,000 *x g* for 1 minute to pellet insoluble debris.

4. Transfer the mixture (supernatant) to a **Zymo-Spin™ IC-XM Column** in a **Collection Tube**. Centrifuge at ≥ 12,000 *x g* for 1 minute. Discard the collection tube with the flow through.

5. Add 200 µl **DNA Pre-Wash** to the spin column in a new **Collection Tube**. Centrifuge at ≥ 12,000 *x g* for 1 minute. Empty the collection tube.

6. Add 700 µl **g-DNA Wash Buffer** to the spin column. Centrifuge at ≥ 12,000 *x g* for 1 minute. Empty the collection tube.

7. Add 200 µl **g-DNA Wash Buffer** to the spin column directly on the matrix. Centrifuge at ≥ 12,000 *x g* for 1 minute. Discard the collection tube with the flow through.

8. Transfer the spin column to a clean microcentrifuge tube. Add ≥ 10 µl **DNA Elution Buffer** or water[3] directly on the matrix. Incubate for 5 minutes at room temperature, then centrifuge at top speed for 1 minute to elute the DNA[4]. The eluted DNA can be used immediately for molecular based applications or stored ≤ -20ºC for future use.

[1] For bone samples, bone powder must be prepared and used for the extraction.
[2] Overnight digestions are possible without affecting the integrity of the DNA.
[3] **DNA Elution Buffer**: 10 mM Tris-HCl, pH 8.5, 0.1 mM EDTA. If water is used, make sure the pH is > 6.0.
[4] The total yield can be improved by eluting the DNA with 60-70 °C **DNA Elution Buffer**. Alternatively, loading the eluate a second time, incubating for 3 minutes at room temperature, and centrifuging again can also increase total yield.

FFPE Tissue[1]

Deparaffinize FFPE Samples:

1. Remove or trim as much paraffin from the sample(s) as possible (≤ 5 mg).

2. Transfer samples to 1.5 ml microcentrifuge tubes. Add 750 µl xylene (not provided) to the samples.

3. Vortex and incubate samples at room temperature for 1 hour with gentle rocking.

4. Centrifuge at 12,000 *x g* for 1 minute and remove the xylene from the sample. Repeat steps 2-4.

5. Wash with 1 ml ethanol (100%), vortex briefly, and incubate for 5 minutes with gentle rocking. Centrifuge at ≥ 12,000 *x g* for 1 minute, discard the supernatant, and repeat.

6. Wash with 1 ml ethanol (95%), vortex briefly, and incubate for 5 minutes with gentle rocking. Centrifuge at ≥ 12,000 *x g* for 1 minute, discard the supernatant, and repeat.

7. Wash with 1 ml ethanol (75%), vortex briefly, and incubate for 5 minutes with gentle rocking. Centrifuge at ≥ 12,000 *x g* for 1 minute, discard the supernatant, and repeat.

8. Wash with 1 ml ddiH_2O, vortex briefly, and incubate for 5 minutes with gentle rocking. Centrifuge at ≥ 12,000 *x g* for 1 minute and remove the water from the sample[2].

DNA Extraction:

1. Prepare the Proteinase K Digestion to the deparaffinized samples in each well as follows[3]:

Water	45 µl
Solid Tissue Buffer (Blue)	45 µl
Proteinase K	10 µl

2. Mix thoroughly or vortex 10-15 seconds and incubate the tube at 55˚C for 12-16 hours. Then incubate the tube at 94˚C for 20 minutes.

[1] The ***Quick*-DNA™ FFPE** (D3067) is specialized for DNA purification from FFPE samples.
[2] It is possible to store samples at -80˚C at this point for later use.
[3] If a ≤ 5 mg tissue sample is not fully submerged in the digestion volume, scale up the digestion to 200 µl while keeping the amount of **Proteinase K** the same.

3. Add 6 volumes **Genomic Binding Buffer** to the tube and mix thoroughly by vortexing for 15 seconds. Centrifuge at ≥ 12,000 *x g* for 1 minute to pellet insoluble debris.

4. Transfer the mixture (supernatant) to a **Zymo-Spin™ IC-XM Column** in a **Collection Tube**. Centrifuge at ≥ 12,000 *x g* for 1 minute. Discard the collection tube with the flow through.

5. Add 200 µl **DNA Pre-Wash Buffer** to the spin column in a new **Collection Tube**. Centrifuge at ≥ 12,000 *x g* for 1 minute. Empty the collection tube.

6. Add 700 µl **g-DNA Wash Buffer** to the spin column. Centrifuge at ≥ 12,000 *x g* for 1 minute. Empty the collection tube.

7. Add 200 µl **g-DNA Wash Buffer** to the spin column directly on the matrix. Centrifuge at ≥ 12,000 *x g* for 1 minute. Discard the collection tube with the flow through.

8. Transfer the spin column to a clean microcentrifuge tube. Add ≥ 10 µl **DNA Elution Buffer** or water[1] directly on the matrix. Incubate for 5 minutes at room temperature, then centrifuge at top speed for 1 minute to elute the DNA[2]. The eluted DNA can be used immediately for molecular based applications or stored ≤ -20ºC for future use.

[1] **DNA Elution Buffer**: 10 mM Tris-HCl, pH 8.5, 0.1 mM EDTA. If water is used, make sure the pH is > 6.0.
[2] The total yield can be improved by eluting the DNA with 60-70 °C **DNA Elution Buffer**. Also, loading the eluate a second time, incubating for 3 minutes at room temperature, and centrifuging again can also increase total yield.

Samples Collected onto Storage Papers/Cards

Rapid purification of inhibitor-free, PCR-quality DNA from blood, saliva, and cells collected onto Guthrie, FTA®, and other storage papers (cards). The procedure is easy; card punches are added directly to a ZR BashingBead™ Lysis Tube (2.0 mm) and thoroughly homogenized by bead beating (e.g. FastPrep®-24, or similar). Following Proteinase K digestion, the DNA is purified using innovative Zymo-Spin™ Technology. Eluted DNA is ideal for PCR, genotyping, etc.

Additional reagents must be purchased separately. For users who plan to process all 50 with this protocol, please see the following ordering information:

Product Name	50 Preps.
ZR BashingBead Lysis Tubes (2.0 mm)[1]	1 x S6003-50
BashingBead Buffer[2]	1 x D6001-3-40
Proteinase K (20 mg)[3]	1 x D3001-2-20
Solid Tissue Buffer (Blue)[4]	2 x D4068-2-6
Genomic Binding Buffer[5]	1 x D4068-3-25

1. Add card samples (punches) to a **ZR BashingBead™ Lysis Tube (2.0 mm)**[1]. Add 400 µl **BashingBead Buffer** to the tube.

2. Secure lysis tube in a bead beater fitted with a 2 ml tube holder assembly and process at maximum speed.

   Processing times may be as little as 40 seconds when using high-speed disrupters (e.g., FastPrep®-24, or similar). See manufacturer's literature for operating instructions.

3. Centrifuge the **ZR BashingBead™ Lysis Tube (2.0 mm)** at ≥ 10,000 x g for 1 minute.

4. To the lysate in the **ZR BashingBead™ Lysis Tube (2.0 mm)**, add:

Proteinase K	40 µl
Solid Tissue Buffer (Blue)	360 µl

5. Mix and then incubate the tube at 55˚C for 10-15 minutes.

6. Centrifuge the **ZR BashingBead™ Lysis Tube (2.0 mm)** at 10,000 x *g* for 1 minute. Transfer 400 µl of supernatant to a microcentrifuge tube.

7. Add 800 µl **Genomic Binding Buffer** to the tube and mix thoroughly.

8. Transfer 600 µl of the mixture to a **Zymo-Spin™ IC-XM Column** in a **Collection Tube**. Centrifuge at ≥ 12,000 *x g* for 1 minute.

9. Discard the flow through from the Collection Tube and repeat Step 8.

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[1] **ZR BashingBead Lysis Tubes (2.0 mm)** - 50 pack: D6003-50
[2] **BashingBead Buffer** - 40 ml: D6001-3-40; 150 ml: D6001-3-150
[3] **Proteinase K Set** - 5 mg: D3001-2-5; 20 mg: D3001-2-20
[4] **Solid Tissue Buffer** - 6 ml: D4068-2-6; 10 ml: D4068-2-10; 22 ml: D4068-2-22
[5] **Genomic Binding Buffer** - 25 ml: D4068-3-25; 45 ml: D4068-3-45; 85 ml: D4068-3-85

10. Add 200 µl **DNA Pre-Wash Buffer** to the spin column in a new Collection Tube. Centrifuge at ≥ 12,000 *x g* for 1 minute. Empty the collection tube.

11. Add 700 µl **g-DNA Wash Buffer** to the spin column. Centrifuge at ≥ 12,000 *x g* for 1 minute. Empty the collection tube.

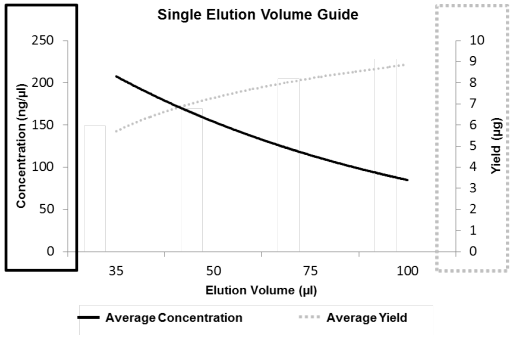
12. Add 200 µl **g-DNA Wash Buffer** to the spin column. Centrifuge at ≥ 12,000 *x g for* 1 minute. Discard the collection tube with the flow through.

13. Transfer the spin column to a clean microcentrifuge tube. Add ≥ 10 µl **DNA Elution Buffer** or water[1] directly on the matrix. Incubate for 5 minutes at room temperature, then centrifuge at maximum speed for 1 minute to elute the DNA[2]. The eluted DNA can be used immediately for molecular based applications or stored ≤ -20ºC for future use.

[1] **DNA Elution Buffer**: 10 mM Tris-HCl, pH 8.5, 0.1 mM EDTA. If water is used, make sure the pH is > 6.0.
[2] The total yield can be improved by eluting the DNA with 60-70 ˚C **DNA Elution Buffer**. Also, loading the eluate a second time, incubating for 3 minutes at room temperature, and centrifuging again can also increase total yield.

Troubleshooting

Problem	Possible Causes and Suggested Solutions
Low DNA Yield	***Incomplete Debris Removal:*** • For solid tissue samples, ensure lysate is centrifuged after digestion to pellet insoluble debris. Transfer the supernatant while avoiding any pelleted debris or lipid layer. ***Incomplete Lysis/Digestion:*** • Ensure Proteinase K digestions are performed at 55˚C as indicated. It is possible to extend digestion times if samples are high in protein. • Mix samples longer after the addition of Genomic Binding Buffer to ensure that the lysate is homogenous. ***Tissue Input:*** • For low DNA-containing tissues (e.g. muscle, etc.) using larger inputs will increase yields (≤ 5mg). • If the lysate does not pass through the column or is extremely viscous, use less input material. Too much tissue can cause cellular debris to overload the column and leech salts into the DNA eluate. ***Procedural Errors:*** • Ensure the proper digestion buffer is used. See the Purification Guide on page 6. • Ensure the correct volume of Genomic Binding Buffer is used. For plasma and serum samples, use 3 volumes of Genomic Binding Buffer. See the Purification Guide on page 6 and the Protocol on page 7. ***Elution Procedures:*** • Ensure the DNA Elution Buffer hydrates the matrix for 5 minutes at room temperature before centrifugation. • To increase yields, heat the DNA Elution Buffer to 60-70˚C before use. You can also load the eluate a second time, incubate at room temperature for 3 minutes, and centrifuge again.

Problem	Possible Causes and Suggested Solutions
Low DNA Yield	***Identifying Proper Elution Volume:***  **The Relationship Between Elution Volume, DNA Yield, and DNA Concentration Using Porcine Whole Blood (Column Format)**. Using a smaller elution volume results in higher concentrations of DNA samples, but with reduced yields. Using a larger elution volume results in higher DNA yields, but at a reduced concentration. Choose an elution volume that best fits your individual application.
DNA Degradation	***DNase Contamination:*** • Check pipettes, pipette tips, microcentrifuge tubes, etc. for DNase contamination and exercise the appropriate precautions during the DNA purification procedure. All reagents and components supplied with the ***Quick*-DNA™ Microprep Plus Kit** are DNase-free. However, DNase contamination can result during the processing of some samples. • If water is used to elute the DNA, ensure that DNase-free water is used. ***Storage:*** • Certain samples are more prone to degradation as a result of the conditions used for storage and transport (e.g. FFPE Tissue).

Problem	Possible Causes and Suggested Solutions
Low DNA Performance	***Procedural Errors:*** • The tip of a column is contaminated with wash buffer flow through. Avoid tilting the column during the wash steps and ensure the tip does not touch the flow through. Empty the collection tube when instructed. • Insufficient centrifugation: Ensure the indicated centrifugation times and speeds are used. Increase the centrifugation time of the final wash step by 1 minute to ensure complete wash buffer removal. ***Incomplete Debris Removal:*** • For solid tissue samples, ensure lysate is centrifuged after digestion to pellet insoluble debris. Ensure pellet is not transferred to the column. ***Tissue Input:*** • Make sure the lysate has passed completely through the matrix before proceeding to the wash steps. • Vortex samples longer after the addition of Genomic Lysis Buffer to ensure that the lysate is homogenous. • If the lysate does not pass through the column or is extremely viscous, use less input material. Too much tissue can cause cellular debris to overload the column and leech salts into the DNA eluate. ***RNA in Eluate:*** • All reagents and components supplied with the ***Quick-DNA™ Microprep Plus Kit*** are designed for RNA removal. Typically if RNA is in the eluate, too much tissue/sample was used. • Ensure the proper amount of Genomic Binding Buffer and corresponding digestion buffer is used. See the Purification Guide on page 6. • Ensure Proteinase K digestions are performed at 55˚C as indicated. • For applications sensitive to trace amounts of RNA, additional RNA removal may be necessary using an RNase A treatment.

Ordering Information

Product Description	Catalog No.	Size
***Quick*-DNA™ Miniprep Plus Kit**	D4068 D4069	50 preps. 200 preps.
***Quick*-DNA™ Microprep Plus Kit**	D4074	50 preps.
***Quick*-DNA™ 96 Plus Kit**	D4070 D4071	2 x 96 preps. 4 x 96 preps.
***Quick*-DNA™ Midiprep Plus Kit**	D4075	25 Preps

Individual Kit Components	Catalog No.	Amount
Proteinase K & Storage Buffer	D3001-2-5 D3001-2-20	5 mg set 20 mg set
BioFluid & Cell Buffer (Red)	D4068-1-12 D4068-1-45	12 ml 45 ml
Solid Tissue Buffer (Blue)	D4068-2-6 D4068-2-22	6 ml 22 ml
Genomic Binding Buffer	D4068-3-25 D4068-3-85	25 ml 85 ml
DNA Pre-Wash Buffer	D3004-5-15 D3004-5-30 D3004-5-50	15 ml 30 ml 50 ml
g-DNA Wash Buffer	D3004-2-50 D3004-2-100 D3004-2-200	50 ml 100 ml 200 ml
DNA Elution Buffer	D3004-4-4 D3004-4-10 D3004-4-50	4 ml 10 ml 50 ml
Zymo-Spin™ IC-XM Columns	C1103-50	50 Columns
Collection Tubes	C1001-50 C1001-500 C1001-1000	50 tubes 500 tubes 1,000 tubes

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EZ DNA Methylation Kits	Size	Catalog No.
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EZ-96 DNA Methylation-Lightning Kit	2x96 Rxns. (Deep-Well) 2x96 Rxns. (Shallow-Well)	D5032 D5033
EZ DNA Methylation-Lightning Automation Kit	96 Rxns.	D5049
EZ-96 DNA Methylation Lightning MagPrep	4 X 96 Rxns. 8 X 96 Rxns.	D5046 D5047

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Library Prep Kits	Size	Catalog No.
Zymo-Seq WGBS Library Kit	24 Preps.	D5465
Pico Methyl-Seq Library Prep Kit	10 Preps. 25 Preps.	D5455 D5456
Zymo-Seq RRBS Library Kit	24 Preps. 48 Preps.	D5460 D5461

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ZymoTaq Polymerase	Size	Catalog No.
ZymoTaq Premix	50 Rxns. 200 Rxns.	E2003 E2004
ZymoTaq DNA Polymerase	50 Rxns. 200 Rxns.	E2001 E2002
ZymoTaq qPCR Premix	50 Rxns. 200 Rxns.	E2054 E2055

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DNA Methylation Standards	Size	Catalog No.
Human Methylated & Non-methylated DNA Set	5 µg/20 µl	D5014
Universal Methylated DNA Standard	Human Mouse	D5011 D5012
Bisulfite-Converted Universal Methylated Human DNA Standard	1 µg/50 µl	D5015
Human Methylated & Non-Methyated (WGA) DNA Set	5 µg/20 µl	D5013

Notes

Notes

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