

RNA Clean & Concentrator™ -100

Clean-up RNA from any sample

Highlights

- Quick, 15-minute clean-up of total RNA (including small/microRNAs) from any enzymatic reaction, aqueous phase following TRIZOL® extraction, in vitro transcription products, etc.
- Ultra-pure RNA is ready for Next-Gen Sequencing, RT-qPCR, etc.

Catalog Numbers:
R1019



Scan with your smart-phone camera to
view the online protocol/video.



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Product Contents

| RNA Clean & Concentrator™-100 | R1019 (25 prep) |
|--|----------------------------|
| RNA Binding Buffer | 100 ml |
| RNA Prep Buffer | 10 ml |
| RNA Wash Buffer (concentrate) ¹ | 6 ml |
| DNase/RNase-Free Water | 10 ml |
| Zymo-Spin™ V-E Columns (w/ reservoir) | 25 |
| Collection Tubes | 50 |
| Instruction Manual | 1 |

Storage Temperature - Store all kit components (i.e., buffers, columns) at room temperature.

Before use:

1 Add 24 ml 100% ethanol (26 ml 95% ethanol) to the 6 ml **RNA Wash Buffer** concentrate.

Specifications

- **Sample Sources** – Enzymatic reactions (e.g., DNase I treated RNA), the aqueous phase following TRIzol®/chloroform or similar¹ extraction, in vitro transcriptions, etc.
- **Size** – Total RNA including small/microRNAs (≥ 17 nt).
- **Purity** – A_{260}/A_{280} & $A_{260}/A_{230} > 1.8$. RNA is ready for Next-Gen Sequencing, RT-qPCR, etc.
- **Binding Capacity** – 1 mg total RNA (**Zymo-Spin™ V-E Column**).
- **Elution Volume** – ≥ 100 μ l **DNase/RNase-Free Water**.
- **Equipment Needed** (user provided) – Vacuum manifold, microcentrifuge.
- **Chemical Tolerance** – $\leq 5\%$ Triton X-100, $\leq 5\%$ Tween-20, $\leq 5\%$ Sarkosyl, $\leq 0.1\%$ SDS. Also compatible with $\leq 90\%$ Sucrose, $\leq 90\%$ Formamide, and $\leq 2\%$ Formaldehyde.

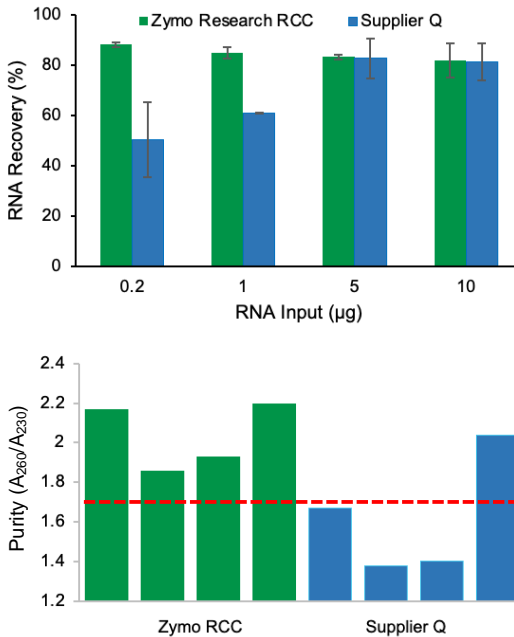
¹ TRI Reagent®, RNAzol®, QIAzol®, TriPure™, TriSure™, and all other acid guanidinium-phenol reagents.

Product Description

The **RNA Clean & Concentrator™-100** kit provides a simple and reliable method for the rapid preparation of up to 1 mg of high-quality, NGS-ready RNA. This 15 minute procedure is based on the use of a unique single-buffer system and Zymo-Spin™ column technology that allows for selective recovery of total RNA (> 17 nt), large RNAs (> 200 nt), and/or small RNAs (17-200 nt).

The procedure is easy: Add binding buffer to your sample, then bind, wash and elute ultra-pure RNA. The RNA can be eluted from the **Zymo-Spin™ V-E Column** in ≥ 100 µl of RNase-free water. The highly concentrated, purified RNA is suitable for all subsequent analyses and molecular manipulations.

Consistent Recovery and Ultra-pure Total RNA



(top) Increasing amounts of RNA was cleaned up using the **RCC™** kit and a Supplier Q kit (n=2). **RCC™** provides higher yields and more consistent recovery when compared to the Supplier Q Kit.

(bottom) RNA was cleaned using the **RCC™** kit and a Supplier Q kit (n=4). RNA purity (measured by A₂₆₀/A₂₃₀) was greater than 1.8 for the **RCC™** kit but not for the Supplier Q kit.

Protocol

The protocol consists of: (I) Buffer Preparation, (II) Sample Preparation and (III) Total RNA Clean-up.

(I) Buffer Preparation

- ✓ Add 24 ml 100% ethanol (26 ml 95% ethanol) to the 6 ml **RNA Wash Buffer** concentrate.

(II) Total RNA Clean-up

- ✓ RNA species ≥ 17 nt will be recovered.
- ✓ Perform all steps at room temperature and centrifugation at 10,000-16,000 x g for 30 seconds, unless specified.
- ✓ For DNA-free RNA (optional), perform **DNase I** treatment before clean-up (page 6).

1. Add 2 volumes **RNA Binding Buffer** to each sample¹ and mix.

Example: Mix 1 ml buffer and 0.5 ml sample.

2. Add an equal volume of ethanol (95-100%) and mix.

Example: Add 1.5 ml ethanol.

3. Transfer the sample to the **Zymo-Spin™ V-E Column w/ Reservoir** mounted on a vacuum manifold and start vacuum¹. Discard the flow-through. Place column into a **Collection Tube**.
4. Add 400 μ l **RNA Prep Buffer** to the column and centrifuge. Discard the flow-through.
5. Add 400 μ l **RNA Wash Buffer** to the column and centrifuge. Discard the flow-through.
6. Add 400 μ l **RNA Wash Buffer** to the column and centrifuge for 1 minute ensure complete removal of the wash buffer. Carefully, transfer the column into a RNase-free tube (not provided).
7. Add ≥ 100 μ l **DNase/RNase-Free Water** directly to the column matrix and centrifuge.

The eluted RNA can be used immediately or stored frozen.

¹ Set vacuum source at ≥ 500 mm Hg.

Appendices

DNase I Treatment

- ✓ For DNA-free RNA, **DNase I** treatment can be performed using **DNase I Set** (E1010; qty: 2) and RNA Wash Buffer (concentrate) (D1003-3-6); materials sold separately.
- ✓ Perform all steps at room temperature and centrifugation at 10,000-16,000 x g for 30 seconds, unless specified.

DNase I treatment before RNA clean-up

For each sample to be treated, prepare 200 µl **DNase I Reaction Mix** in an RNase-free tube (not provided) and mix by gentle inversion. Then incubate at room temperature (20-30°C) for 15 minutes and proceed with the RNA Clean-up protocol, page 5.

DNase I Reaction Mix

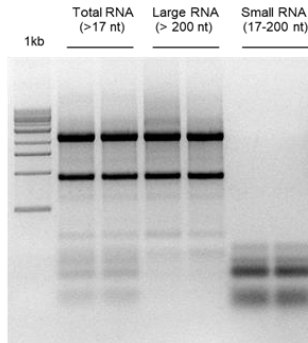
| | |
|---|--------|
| RNA sample (≤ 250 µg; volume adjusted with water or TE buffer) | 160 µl |
| DNase I (reconstituted; 1 U/ul) ^{1,2} | 20 µl |
| DNA Digestion Buffer | 20 µl |

1 Unit definition – one unit increases the absorbance of a high molecular weight DNA solution at a rate of 0.001 A260 units/ml of reaction mixture at 25°C.

2 Reconstitute each vial of lyophilized **DNase I** (#E1009-A; 250 U) with 275 µl **DNase/RNase-Free Water** and mix by gentle inversion. Store frozen aliquots.

Purification of Small and Large RNAs into Separate Fractions

- ✓ Perform all steps at room temperature and centrifugation steps at 10,000-16,000 x g for 30 seconds, unless specified.
 - ✓ This protocol requires two column per prep.
1. Prepare adjusted **RNA Binding Buffer** (as needed). Mix an equal volume of buffer and ethanol (95-100%).
Example: Mix 500 µl buffer and 500 µl ethanol.
 2. Add 2 volumes of the adjusted buffer to the sample¹ and mix.
Example: Mix 1 ml adjusted buffer and 0.5 ml sample.
 3. Transfer the mixture to the **Zymo-Spin™ Column**² and centrifuge.
Save the flow-through!
 4. **Small RNAs (17-200 nt) are in the flow-through**
 - a. Add 1 volume ethanol and mix.
Example: Add 1.5 ml ethanol to 1.5 ml sample.
 - b. Transfer the mixture to a **new column** and centrifuge. Discard the flow-through.
 - c. Proceed with the RNA Clean-up protocol, page 5, step 4.
 4. **Large RNAs (> 200 nt) are retained in the column**
 - a. Proceed with the RNA Clean-up protocol, page 5, step 4.



RNA Clean & Concentrator™ allows for clean-up of total RNA (> 17 nt), large RNAs (> 200 nt), and/or small RNAs (17-200 nt).

1 To minimize pipetting error, adjust the sample volume to 50 µl (minimum).
2 To process samples >700 µl, **Zymo-Spin™** columns may be reloaded.

RNA clean-up from aqueous phase after TRIzol®/chloroform extraction

Following TRIzol®/chloroform or similar* extraction, carefully transfer the upper aqueous phase into an RNase-free tube (not provided). Add 1 volume of ethanol (95-100%) to 1 volume of aqueous phase¹ (1:1) and mix well. Then proceed with the RNA Clean-up protocol, page 5, step 3.

RNA clean-up from samples in DNA/RNA Shield™

- ✓ Perform all steps at room temperature and centrifugation steps at 10,000-16,000 x g for 30 seconds, unless specified.
- 1. If frozen, thaw samples to room temperature (20-30°C) and centrifuge debris (if any). Transfer the cleared sample into an RNase-free tube (not provided).
- 2. Add 1 volume of ethanol (95-100%) to 1 volume of the **DNA/RNA Shield™** sample¹ and mix well.
Example: 50 µl buffer and 50 µl sample.
- 3. Continue with the RNA Clean-up protocol, page 5, step 3.

cDNA Clean-Up following Reverse Transcription (RT)

The **RNA Clean & Concentrator** can be used to effectively clean and concentrate first-strand cDNA following reverse transcription (RT) and hydrolysis. The **RNA Binding Buffer** will neutralize the hydrolysis reaction and the recovered cDNA may be used directly for microarray analysis, etc.

Hydrolysis Reaction: To each 30-50 µl RT reaction, add 10 µl 0.5 M EDTA and 10 µl 1 M NaOH. Then mix and incubate at 65°C for 15 minutes. Proceed to the Total RNA Clean-Up protocol, page 5.

* TRI Reagent®, RNAzol®, QIAzol®, TriPure™, TriSure™, and all other acid guanidinium-phenol reagents.
1 To minimize pipetting error, adjust the sample volume to 50 µl (minimum).

Ordering Information

| Product Description | Catalog No. | Size |
|-------------------------------|-------------|----------|
| RNA Clean & Concentrator™-100 | R1019 | 25preps. |

| Individual Kit Components | Catalog No. | Amount |
|--|-------------|--------|
| RNA Binding Buffer | R1013-2-50 | 50 ml |
| | R1013-2-100 | 100 ml |
| RNA Prep Buffer | R1060-2-25 | 25 ml |
| | R1060-2-100 | 100 ml |
| RNA Wash Buffer (concentrate) | R1003-3-6 | 6 ml |
| | R1003-3-24 | 24 ml |
| Zymo-Spin™ V-E Columns (w/ reservoir) | C1029-25 | 25 |
| Collection Tubes | C1001-50 | 50 |
| | C1001-500 | 500 |
| DNase/RNase-Free Water | W1001-6 | 6 ml |
| | W1001-10 | 10 ml |
| DNase I Set (250 U DNase I (lyophilized) supplied with DNA Digestion Buffer, 4 ml) | E1010 | 1 |

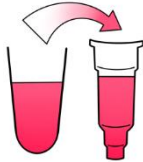
Complete Your Workflow

- ✓ For tough-to-lyse samples in TRIzol, use ZR BashingBead Lysis Tubes:

ZR BashingBead Lysis Tubes

| | |
|---------------------------|--------------------------------|
| 2.0 mm beads #S6003 | For plant/animal tissue |
| 0.1 + 0.5 mm beads #S6012 | For microbes |
| 0.1 + 2.0 mm beads #S6014 | For microbes in tissue/insects |

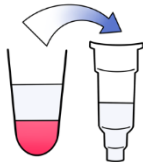
- ✓ The only **direct**, high-throughput and automatable RNA purification from sample lysates in TRIzol (DNase I Set included with all formats):



Direct-zol RNA kits

| | |
|----------------------------|---|
| Microprep #R2060-R2063 | From 1 cell and up |
| Miniprep #R2050-R2053 | Up to 50 ug RNA |
| Miniprep Plus #R2070-R2073 | Up to 100 ug RNA |
| 96-well #R2054-R2057 | Spin-plate |
| MagBeads #R2100-R2105 | Automatable (Tecan, Hamilton, Kingfisher, etc.) |

- ✓ For RNA clean-up (purification) from the aqueous phase (e.g., TRIzol, TRI Reagent or similar) or from any enzymatic reaction (e.g., DNase I treated RNA):



RNA Clean & Concentrator kit

| | |
|--------------|----------------------|
| #R1013-R1014 | DNase I Set included |
|--------------|----------------------|





- ✓ For NGS:

Zymo-Seq RiboFree Total RNA Library Prep kit

| | |
|--------|----------|
| #R3000 | 12 preps |
| #R3003 | 96 preps |

Format Compatibility

To adjust binding capacity, simply replace the provided columns with the formats indicated below and follow their respective kit protocol.

| Format | Zymo-Spin™ I & IC | Zymo-Spin™ II & IICR | Zymo-Spin™ V-E | Zymo-Spin™ I-96 Plate |
|------------------|---|---|---|---|
| Item Image |  |  |  |  |
| Kit Name | RCC™-5 | RCC™-25 | RCC™-100 | RCC™-96 |
| Capacity | 10 µg / prep. | 50 µg / prep. | 1 mg / prep. | 10 µg / prep. |
| Elution Vol. | ≥ 6 µl | ≥ 25 µl | ≥ 100 µl | ≥ 10 µl |
| Column Cat. Nos. | C1003-50 , C1004-50 | C1008-50 , C1078-50 | C1024-25 , C1029-25 | C2004 |



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