



ZYMO RESEARCH

RNA
Purification
Made Simple

Quick-RNA™ Plant Miniprep

RNA from any tough-to-lyse sample

Highlights

- Quick, spin-column purification of total RNA (including small/microRNAs) from a wide variety of plant samples (e.g., leaves, stems, buds, flowers, fruit, seeds, etc.)
- ZR BashingBead™ Lysis Tubes are ultra-high density, fracture resistant, chemically inert ceramic beads and used for the robust homogenization of any tough-to-lyse sample.
- RNA is ready for Next-Gen Sequencing, RT/qPCR and any downstream application, etc.

Catalog Numbers:

R2024



Scan with your smart-phone camera to view the online protocol/video.



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Product Contents

Quick-RNA™ Plant Miniprep	R2024 (50 prep)
RNA Lysis Buffer	50 ml
RNA Prep Buffer	25 ml
RNA Wash Buffer ¹ (concentrate)	24 ml
DNase/RNase-Free Water	4 ml
Prep Solution	30 ml
ZR BashingBead™ Lysis Tubes (2.0 mm)	50
Zymo-Spin™ IIIGC Columns	50
Zymo-Spin™ IICR Columns	50
Zymo-Spin™ III-HRC Filters	50
Collection Tubes	100
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Storage Temperature - Store all kit components (i.e., buffers, columns) at room temperature.
Before use:

1 Add 96 ml 100% ethanol (104 ml 95% ethanol) to the 24 ml **RNA Wash Buffer** concentrate.

Specifications

- **Sample Sources** – Up to 150 mg plant tissue (e.g., leaves, stems, buds, flowers, fruit, seeds, etc.)
- **Size** – Total RNA including small/microRNAs (≥ 17 nt).
- **Purity** – A_{260}/A_{280} & $A_{260}/A_{230} > 1.8$. RNA is ready for Next-Gen Sequencing, RT/qPCR, etc.
- **Binding Capacity** – Zymo-Spin™ IICR Column yield up to 50 μ g RNA.
- **Compatibility** – For samples stored in DNA/RNA Shield™, see page 6. DNA/RNA Shield™ lyses cells, inactivates nucleases and infectious agents (e.g., virus, pathogens) and is ideal for safe sample storage and transport at ambient temperatures.
- **Elution Volume** – ≥ 25 μ l DNase/RNase-Free Water.
- **Equipment Needed (user provided)** – Microcentrifuge, vortex, and a high-speed homogenizer/cell disruptor or bead beater (e.g., MP Bio FastPrep-24, Bertin Precellys, etc.) (recommended).

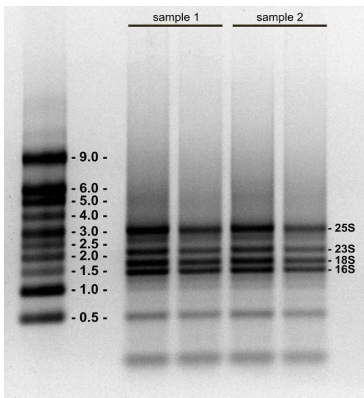
Product Description

The **Quick-RNA™ Plant Miniprep** kit provides for rapid (10-minute) isolation of RNA from various plant samples (e.g. leaves, stems, buds, flowers, fruit, seeds etc.). For purification of total RNA (including small/microRNAs) up to 50 µg.

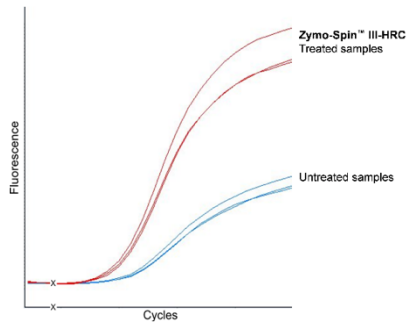
The kit includes unique technology such as the **ZR BashingBead™ Lysis Tubes** and features a specially formulated **RNA Lysis Buffer**. The **Zymo-Spin™ IIICG Column** allows for high-capacity DNA elimination and the subsequent **Zymo-Spin™ IICR Column** efficiently adsorbs total RNA.

The RNA is washed and then eluted with **DNase/RNase-Free Water**. For inhibitor removal, the eluted RNA can be treated by running the sample through the **Zymo-Spin™ III-HRC Filters**. RNA is eluted in as little as 6 µl and is suitable for subsequent procedures including RT-qPCR.

Efficient Recovery of RNA from any Plant Sample



Isolation of total RNA from 10 mg of a fresh leaf (*Nicotiana* sp.) using the **Quick-RNA™ Plant Miniprep** kit. Leaves were minced and processed with a **FastPrep®-24** device (MP Biomedicals). Samples 1 and 2 were loaded in 2x and 1x volume aliquots, respectively, and resolved in a 1% (w/v) nondenaturing agarose gel. RNA Millenium™ Markers (Ambion) were used as size standards.



Nicotiana sp. leaf samples were spiked with humic acid (Sigma) at a final $Ab_{230nm} = 0.2$. Total RNA was isolated with and without the use of the **Zymo-Spin™ III-HRC Filter**. RT-PCR performed with a **LightCycler™ 480** (Roche) showed an increase in fluorescence signal and detected an early amplification initiation for the **Zymo-Spin™ III-HRC** treated samples compared to the non-treated samples ($c_p = [30$ vs. $31]$, respectively).

Protocol

The protocol consists of: (I) Buffer Preparation and (II) Total RNA Purification

(I) Buffer Preparation

- ✓ Add 96 ml 100% ethanol (104 ml 95% ethanol) to the 24 ml **RNA Wash Buffer**.

(II) Total RNA Purification

- ✓ Perform all steps at room temperature and centrifugation at 10,000-16,000 x g for 30 seconds, unless specified.
1. Transfer up to 150 mg fresh or frozen, finely minced (pre-cut) plant tissue into a **ZR BashingBead Lysis Tube** (2.0 mm) and add 800 μ l **RNA Lysis Buffer**.
 2. Secure tube in a bead beater fitted with a 2 ml tube hold assembly and process¹.
 3. Centrifuge the tube for 1 minute to pellet debris.
 4. Transfer up to 400 μ l of the cleared supernatant into a **Zymo-Spin™ IICG Column²** in a **Collection Tube** and centrifuge. Save the flow-through!
 5. To the flow-through, add an equal volume ethanol (95-100%) (1:1) and mix well.
 6. Transfer the mixture into a **Zymo-Spin™ IICR Column²** in a **Collection Tube** and centrifuge. Discard the flow-through.
Optional: At this point, DNase I treatment can be performed. See page 6.
 7. Add 400 μ l **RNA Prep Buffer** to the column and centrifuge. Discard the flow-through.
 8. Add 700 μ l **RNA Wash Buffer** to the column and centrifuge. Discard the flow-through.
 9. Add 400 μ l **RNA Wash Buffer** and centrifuge the column for 1 minute to ensure complete removal of the wash buffer. Then carefully, transfer the column into a nuclease-free tube (not provided).
 10. Add 50 μ l **DNase/RNase-Free Water** directly to the column matrix and centrifuge.
Alternatively, for highly concentrated RNA use ≥ 25 μ l elution.
 11. Place a **Zymo-Spin™ III-HRC Filter** in a new **Collection Tube** and add 600 μ l **Prep Solution**. Centrifuge at 8,000 x g for 3 minutes.
 12. Transfer the eluted RNA (step 10) into the prepared filter in an RNase-free tube (not provided) and centrifuge at 16,000 x g for 3 minutes.

The eluted RNA can be used immediately or stored frozen.

¹ Process with high-speed homogenizer (e.g., MP Bio FastPrep-24, Bertin Precellys) for 30-60 seconds or low-speed homogenizer (e.g., Disruptor Genie) for 3-5 minutes.

² To process samples > 700 μ l, columns may be reloaded.

Appendices

Samples stabilized and stored in DNA/RNA Shield™

1. If frozen, thaw homogenized sample in **DNA/RNA Shield™** to room temperature (20-30°C). Then transfer the entire sample to a **ZR BashingBead Lysis Tube** and bring the total volume of **DNA/RNA Shield™** to 800 µl.
2. Secure tube in a bead beater fitted with a 2 ml tube hold assembly and process².
3. Centrifuge the tube for 1 minute to pellet debris.
4. Transfer the cleared supernatant into a new tube (not provided).
5. Add 1 volume of **RNA Lysis Buffer** to the supernatant (1:1) and mix.
6. Transfer up to 400 µl of the mixture into a **Zymo-Spin™ IIICG Column³** in a **Collection Tube** and centrifuge. Save the flow-through!
7. Proceed to Total RNA Purification, page 5, step 5.

DNase I Treatment (in-column)

- ✓ Perform DNase I treatment with DNase I Set (#E1010) and RNA Wash Buffer (concentrate; #R1003-3-6); available separately.
 - ✓ Perform all steps at room temperature and centrifugation at 10,000-16,000 x g for 30 seconds, unless specified.
1. Following RNA binding step (page 5, step 6), add 400 µl **RNA Wash Buffer** to the column, centrifuge and discard the flow-through.
 2. For each sample to be treated, prepare **DNase I Reaction Mix** (see table below) in an RNase-free tube (not provided) and mix by gentle inversion. Then add 80 µl directly into column matrix and incubate at room temperature (20-30°C) for 15 minutes. Proceed with the purification (page 5, step 7).

DNase I Reaction Mix

DNase I (reconstituted; 1 U/ul) ¹	5 µl
DNA Digestion Buffer	75 µl

1 Prior to use, reconstitute the lyophilized **DNase I** with 275 µl DNase/RNase-Free Water. Mix by gentle inversion and store frozen aliquots. * Unit definition – one unit increases the absorbance of a high molecular weight DNA solution at a rate of 0.001 A₂₆₀ units/ml of reaction mixture at 25°C.

2 Process with high-speed homogenizer (e.g., MP Bio FastPrep-24, Bertin Precellys) for 30-60 seconds or low-speed homogenizer (e.g., Disruptor Genie) for 3-5 minutes.

3 To process samples > 700 µl, columns may be reloaded.

Ordering Information

Product Description	Catalog No.	Size
Quick-RNA™ Plant Miniprep	R2024	50 preps.

Individual Kit Components	Catalog No.	Amount
RNA Lysis Buffer	R1060-1-50 R1060-1-100	50 ml 100 ml
RNA Prep Buffer	R1060-2-25 R1060-2-100	25 ml 100 ml
RNA Wash Buffer (concentrate)	R1003-3-24 R1003-3-48	24 ml 48 ml
DNase/RNase-Free Water	W1001-10 W1001-30	10 ml 30 ml
DNase I Set (lyophilized) (250 U supplied with DNA Digestion Buffer, 4 ml)	E1010	1 set
Zymo-Spin™ IICG Columns	C1006-50-G	50
Zymo-Spin™ IICR Columns	C1078-50	50
Zymo-Spin™ III-HRC Filters	C1058-50	50
Collection Tubes	C1001-50	50
OneStep PCR Inhibitor Removal Kit	D6030	50
Prep Solution	D6035-1-30	30 ml
ZR BashingBead™ Lysis Tubes (2.0 mm)	S6003-50	50

Complete Your Workflow

- ✓ For tough-to-lyse samples, use ZR BashingBead Lysis Tubes:

ZR BashingBead Lysis Tubes	
2.0 mm beads #S6003	Plant/animal tissue
0.1 + 0.5 mm beads #S6012	Microbes
0.1 + 2.0 mm beads #S6014	Microbes in tissue/insects

- ✓ For isolation of RNA from any sample:

Quick-RNA kits	
Miniprep Plus #R1057/R1058	$\leq 10^7$ cells, ≤ 50 mg tissue
MagBeads #R2132/R2133	Automatable (Tecan, Hamilton, Kingfisher, etc.)

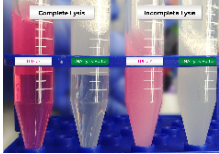
- ✓ For clean-up (purification) and concentration of any RNA sample. (e.g., from the aqueous phase of TRIzol[®] extractions) or from any enzymatic reaction (e.g., DNase I treated RNA):

RNA Clean & Concentrator kits	
Microprep #R1013-R1014	DNase I Set included
MagBeads #R1082	Automatable (Tecan, Hamilton, Kingfisher, etc.)

- ✓ For NGS:

Zymo-Seq RiboFree Total RNA Library Prep kit	
#R3000	12 preps
#R3003	96 preps

Troubleshooting Guide

Problem	Possible Causes and Suggested Solutions
<p>Precipitation, viscous lysate</p>	<p>Incomplete lysis and/or high-mass input:</p> <ul style="list-style-type: none"> - If precipitation occurs (upon adding ethanol to the lysate) or if the lysate is extremely viscous, increase the volume of DNA/RNA Shield™ and/or RNA Lysis Buffer to ensure complete lysis and homogenization until lysate is transparent (see image). 
<p>Low purity (A_{260}/A_{230} nm, A_{260}/A_{280} nm)</p>	<p>Sample handling:</p> <ul style="list-style-type: none"> - Ethanol and/or salt contamination. After centrifugation steps, carefully remove the column from the collection tube to prevent buffer carryover. Alternatively, blot emptied collection tube with a tissue or towel. - Make sure lysate and wash buffers have passed completely through the matrix of the column. This may require centrifuging at a higher speed and/or longer time. <p>Incomplete lysis and/or cellular debris:</p> <ul style="list-style-type: none"> - Increase the volume DNA/RNA Shield™ and/or RNA Lysis Buffer (proportionally) to ensure complete lysis and homogenization. Be sure to centrifuge any cellular debris and then process the cleared lysate.
<p>Low yield</p>	<p>Sample input:</p> <ul style="list-style-type: none"> - Too much input or incomplete lysis/homogenization can cause cellular debris to clog or overload the column and result in compromised nucleic acid recovery. Use less input material and/or increase the volume DNA/RNA Shield™ and/or RNA Lysis Buffer. <p>High-protein content:</p> <ul style="list-style-type: none"> - Perform Proteinase K treatment to the sample prior to purification. See appropriate sample preparation protocol.
<p>DNA contamination</p>	<p>To remove DNA:</p> <ul style="list-style-type: none"> - Perform in-column DNase I treatment or perform DNase I treatment post-purification, then re-purify the treated sample. - For future preps, increase the volume of DNA/RNA Shield™ and/or RNA Lysis Buffer to ensure complete lysis and homogenization of the sample.
<p>RNA degradation</p>	<p>To prevent RNA degradation:</p> <ul style="list-style-type: none"> - Immediately collect and lyse fresh sample into DNA/RNA Shield™ and/or RNA Lysis Buffer ensure stability. Homogenized samples can be stored frozen for later processing.

For technical assistance, please contact 1-888-882-9682 or email tech@zymoresearch.com



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This product is for research use only and should only be used by trained professionals. It is not for use in diagnostic procedures. Some reagents included with this kit are irritants. Wear protective gloves and eye protection. Follow the safety guidelines and rules enacted by your research institution or facility.

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